

Lipid extraction of microalgae and hydrothermal carbonization of the residual algal biomass for sustainable hydrochar production

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1. Introduction and Objectives

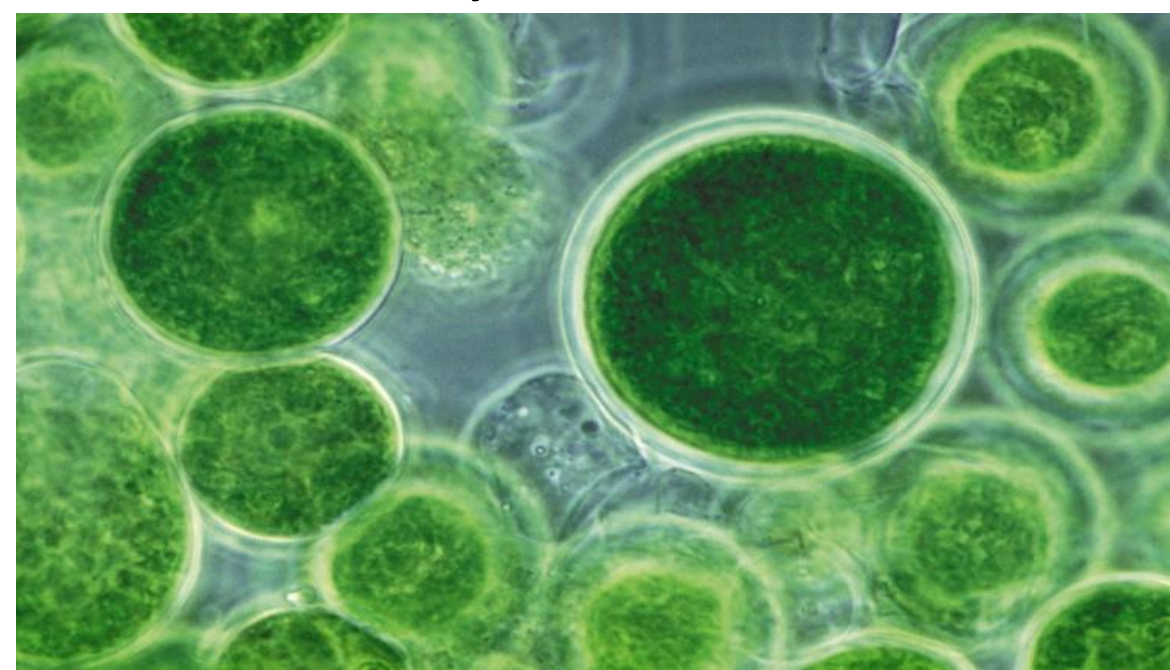
- To enhance the economics of algae technologies, value-added coproducts need to complement lipid production.
- Two routes for lipid extraction either “dried route” or “wet route”. ⁽¹⁾
- For commercial development, cell disruption and lipid extraction from wet biomass need to be scalable and sustainable.
- The solid phase after lipid extraction is termed lipid extracted algae (LEA).
- Hydrothermal Carbonization (HTC) is a process where algal biomass is converted into a solid carbonaceous material called hydrochar. ^(2,3)

Table 1:Energy requirements for different steps in algal biodiesel production . ⁽⁴⁾

Biodiesel production step (basis: 1kg of algal biodiesel)	Dry algal biomass (MJ)	Wet algal biomass (MJ)
Algae cultivation	7.5	10.6
Drying	90.3	0
Extraction	8.6	30.8
Oil transesterification	0.9	0.9
Total	107.3	42.3

Objectives of this study:

- Test cell disruption methods to identify the conditions for maximum lipid yield from *Nannochloropsis oculata*.
- Compare lipid yield among disruption methods.
- Extract lipids to obtain LEA.
- Apply HTC process to LEA in order to obtain hydrochar.
- Evaluate the hydrochar yield under various HTC experiments.
- Characterize hydrochar.



Hydrochar valorization: ⁽⁵⁾

- Soil amendment
- Water treatment
- Solid fuel
- Capacitors
- Low cost adsorbents

2.Materials and experimental conditions

Materials:

- Algae species: Microalga *Nannochloropsis oculata* cultivated in outdoor novel horizontal bioreactor (HBR), harvested and washed with 0.9% salt water.
- A mixture of chloroform/methanol (2:1 % v/v) was used for the lipid extraction.
- LEA obtained after lipid extraction and oven dried to 15% moisture.

Methods & conditions:

- Cell disruption methods
 - Hydrothermal sulfuric acid method ⁽⁶⁾

Variables: a) **Temperature:** 105-125 °C, b) **Residence time:** 10-40 minutes and c) **Acid concentration:** 0.5-3%

- Liquid nitrogen method ⁽⁷⁾
- Mechanical method ⁽¹⁾

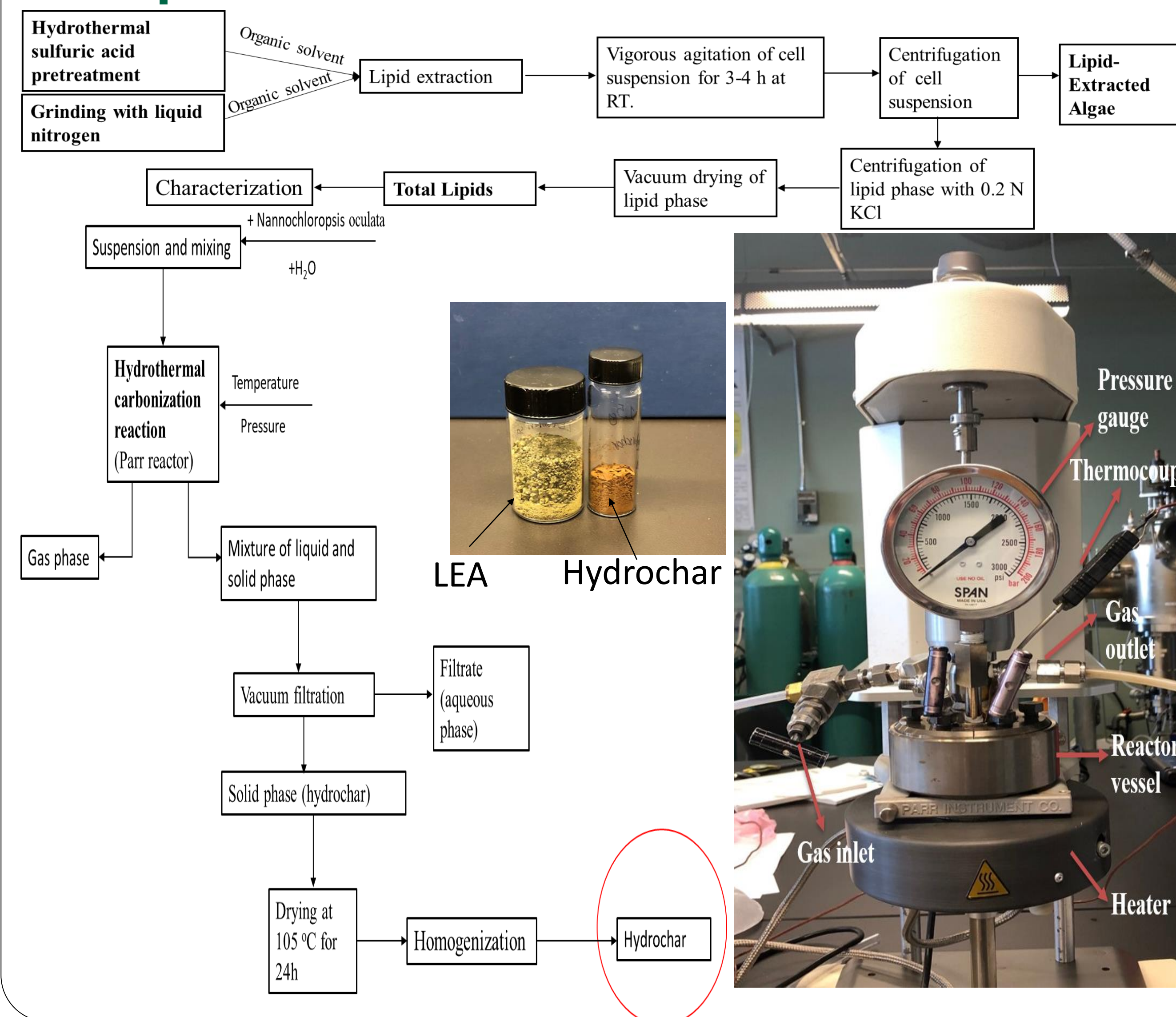
- Hydrothermal carbonization (HTC) ⁽⁸⁾

Variables: a) **Temperature:** 175, 190, 205 & 220 °C, b) **Residence time:** 2h ,and c)**Solid loading:** 8 wt%

$$\text{Hydrochar Yield \%} = \left(\frac{\text{Hydrochar mass}}{\text{LEA mass}} \right) * 100\%$$

$$\text{Lipid yield\%} = \left(\frac{\text{Mass of extracted lipids}}{\text{Total mass of dried algae}} \right) * 100\%$$

3. Experimental Procedure

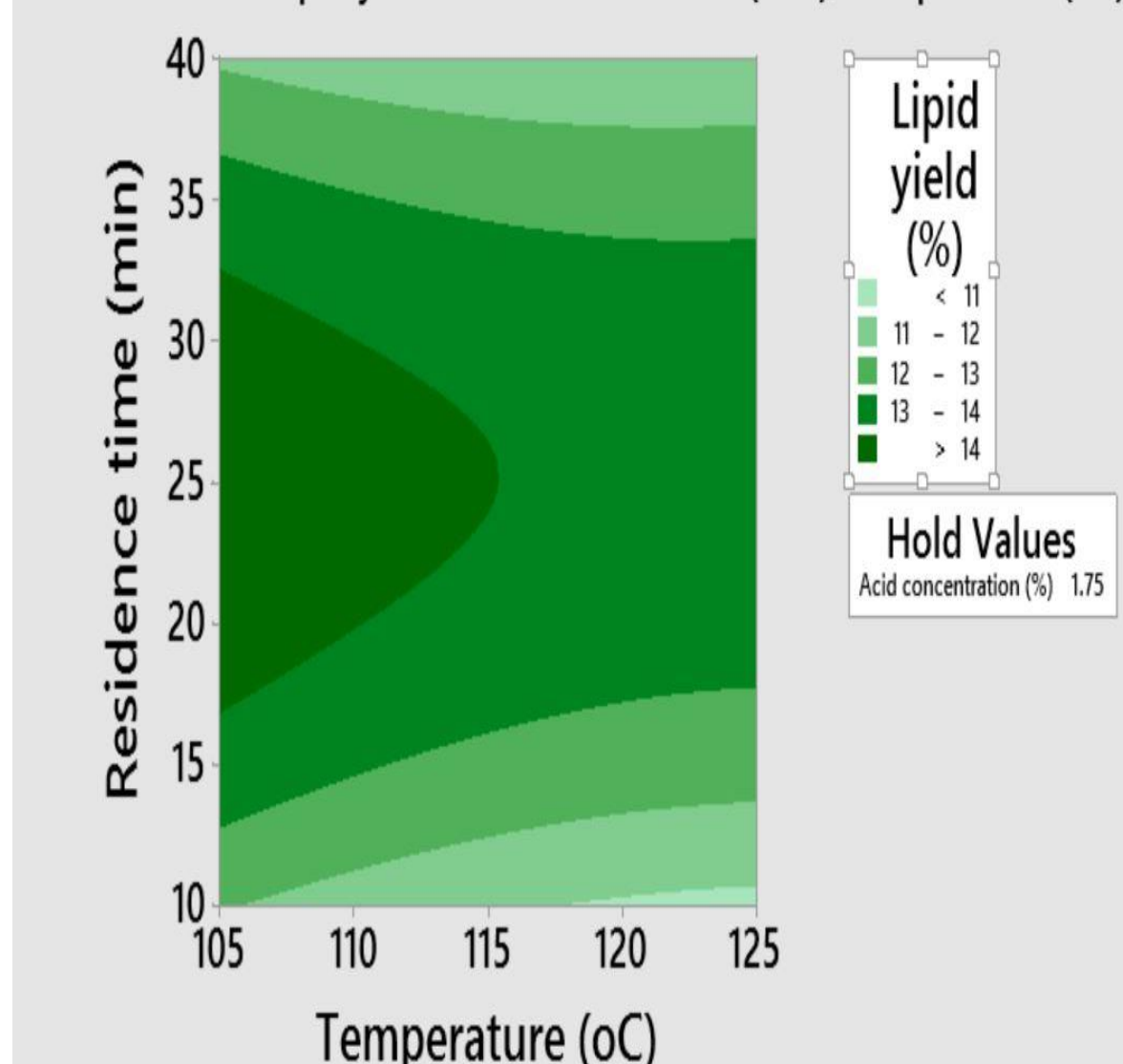


4. Results and Discussion

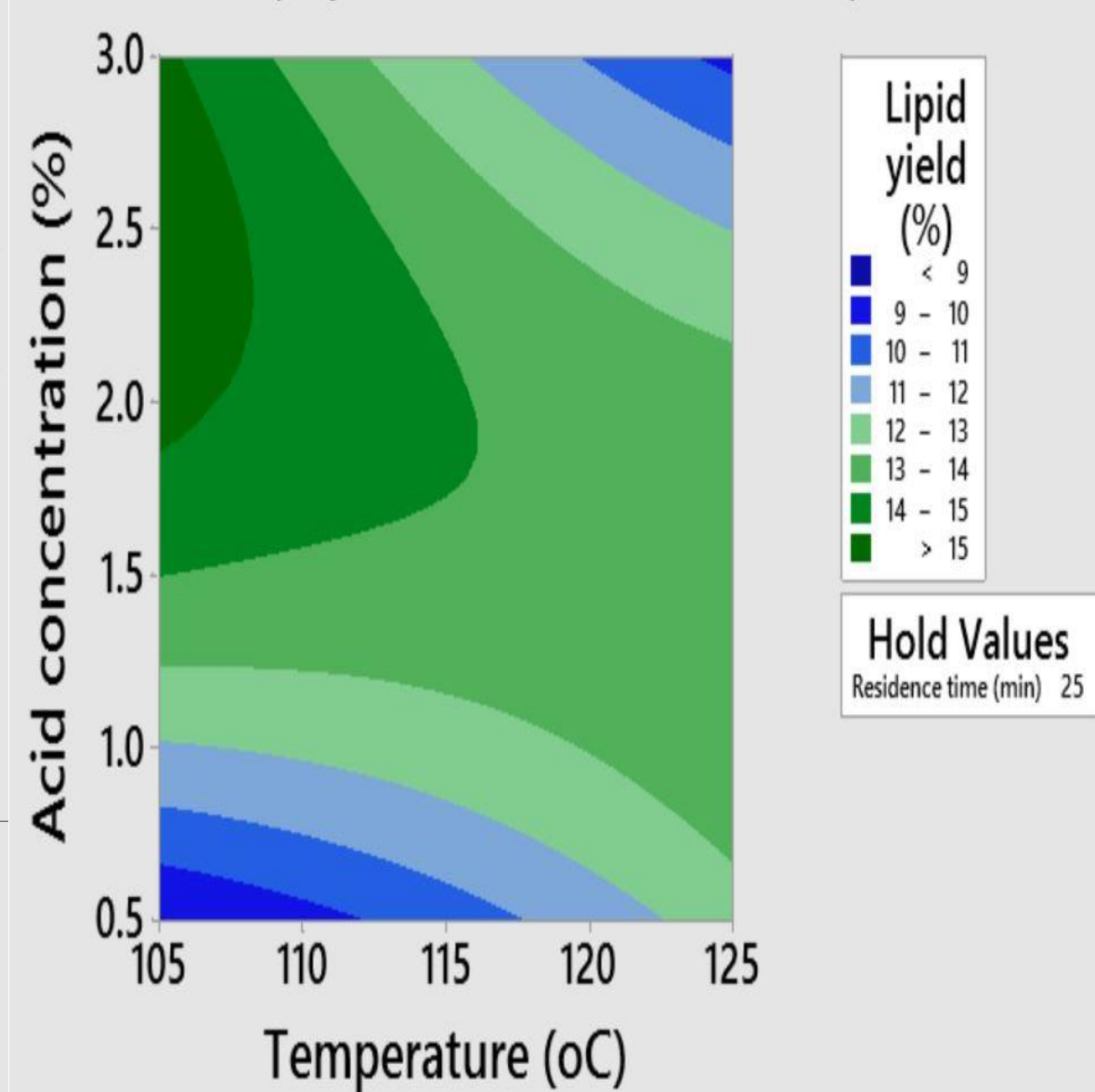
Sulfuric acid treatment statistical optimization

$Y = 1 - 0.34 * X_1 + 0.655 * X_2 + 30.18 * X_3 + 0.00254 * X_1^2 - 0.01242 * X_2^2 - 1.707 * X_3^2 + 0.00124 * X_1 * X_2 - 0.1831 * X_1 * X_3 - 0.0984 * X_2 * X_3$
Where: Y=lipid yield(%), X_1 =Temperature, X_2 = Time and X_3 = Acid concentration (%)

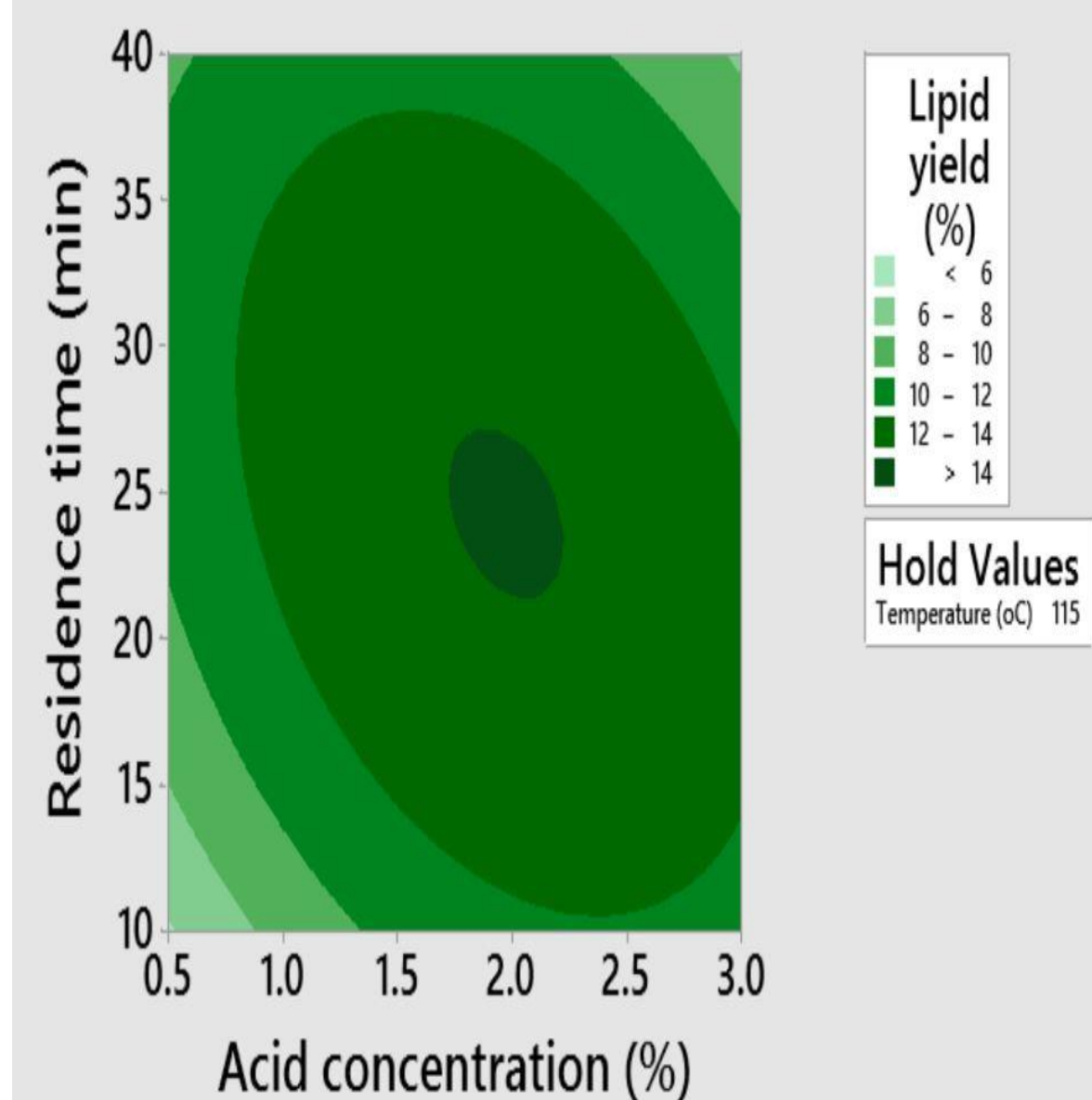
Contour Plot of Lipid yield vs Residence time (min), Temperature (oC)



Contour Plot of Lipid yield vs Acid concentration (%), Temperature (oC)



Contour Plot of Lipid yield vs Residence time (min), Acid concentrati

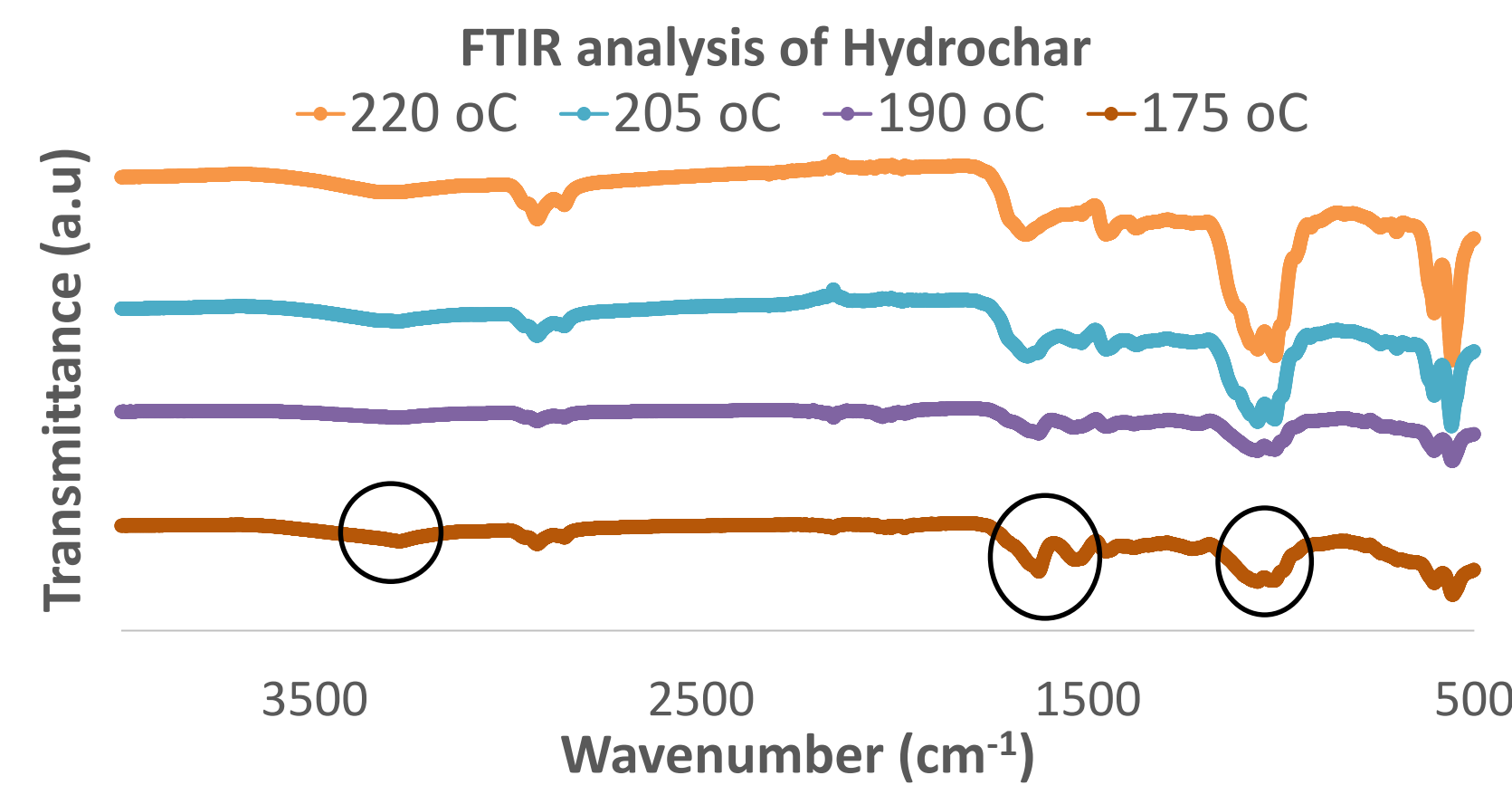
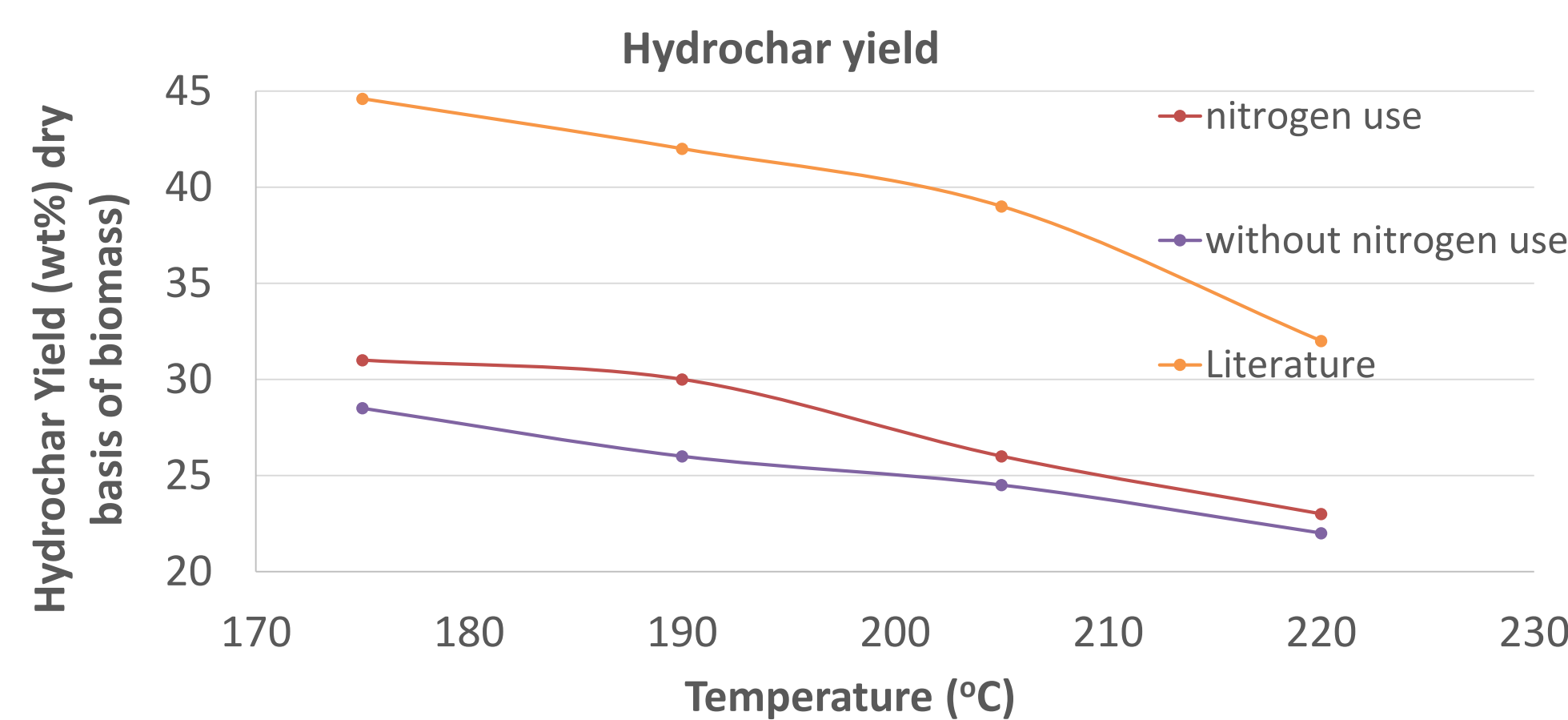


Cell disruption methods comparison

Method	Lipid yield (%)
Hydrothermal sulfuric acid	18.28
Liquid nitrogen	19.84
Mechanical	11.1

Hydrochar Yield, Surface Area, and Pore size

Temperature (°C)	Hydrochar Yield % (Nitrogen Use)	Surface Area (m²/g)	Pore Size (nm)	Hydrochar Yield % (without Nitrogen)	Hydrochar Yield % Literature ^(2,8)
175	31	5.26	7.22	28.5	44.6
190	30	9.2	7.02	27	42
205	26	14.31	6.80	26	39
220	23	18.9	6.20	25.5	32



- The hydrochar yield was 23-31% and decreased as temperature increased.
- The surface area of hydrochar was 5.26-18.9 m²/g.
- Functional groups :

- O-H stretching vibration in hydroxyl and carbonyl groups
- C=O stretching vibration of carbonyl group conjugated with aromatic ring
- C=C stretching of the aromatic ring
- Aromatic C-H bending vibration

5. Summary, Conclusions, and Future Prospects

- Sulfuric acid is a potential catalyst which leads to lipid extraction from *N. oculata*.
- The maximum lipid yield with sulfuric acid treatment is comparable with liquid nitrogen treatment.
- The maximum lipid yield was achieved around the central point based on the RSM model.
- HTC is a promising treatment to produce hydrochar from algal biomass.
- During HTC experiments, in the temperature range 175-220 °C, hydrochar was successfully produced.
- The autogenic pressure during the HTC experiments was 8- 25 bar depending on reaction temperature.
- The highest hydrochar yield was observed at lower temperatures. Hence, lower temperatures enhance the solid formation of the HTC reaction.
- The surface area of hydrochar increased at higher temperatures.
- The functional groups on the surfaces of hydrochars, which was determined by FTIR analysis, are in agreement with the ones reported in the literature. ^(2,3,8)

Future Prospects:

- Use a different lipid extraction mixture, which will be more environmentally friendly.
- Use a different treatment method with lower energy requirements.
- Investigate the influence of solids loading, residence time, and catalyst use on hydrochar yield and quality.
- Characterize the hydrochar and investigate potential applications.
- Optimize the experimental procedure in order to maximize hydrochar yield.

6. Acknowledgements

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7. References

- Ghaly, M. (2015). "Microalgae Oil Extraction Pre-treatment Methods: Critical Review and Comparative Analysis." *Journal of Fundamentals of Renewable Energy and Applications* **05**.
- Heilmann, S. M., H. T. Davis, L. R. Jader, P. A. Lefebvre, M. J. Sadowsky, F. J. Schendel, M. G. von Keitz and K. J. Valentas (2010). "Hydrothermal carbonization of microalgae." *Biomass and Bioenergy* **34**(6): 875-882.
- Lee, J., K. Lee, D. Sohn, Y. M. Kim and K. Y. Park (2018). "Hydrothermal carbonization of lipid extracted algae for hydrochar production and feasibility of using hydrochar as a solid fuel." *Energy* **153**: 913-920.
- Martinez-Guerra, E. and V. G. Gude (2016). "Energy aspects of microalgal biodiesel production." *AIMS Energy* **4**: 347-362.
- Zhang, S., X. Zhu, S. Zhou, H. Shang, J. Luo and D. C. W. Tsang (2019). Chapter 15 - Hydrothermal Carbonization for Hydrochar Production and Its Application. *Biochar from Biomass and Waste*. Y. S. Ok, D. C. W. Tsang, N. Bolan and J. M. Novak, Elsevier: 275-294.
- Lee, I., J.-Y. Park, S.-A. Choi, Y.-K. Oh and J.-I. Han (2014). "Hydrothermal nitric acid treatment for effectual lipid extraction from wet microalgae biomass." *Bioresource technology* **172C**: 138-142.
- Arora, N., A. Patel, P. Pruthi and V. Pruthi (2016). "Synergistic dynamics of nitrogen and phosphorous influences lipid productivity in *Chlorella minutissima* for biodiesel production." *Bioresource Technology* **213**.
- Amber, B., J. Umakanta, S. K. Hoekman and L. Joel (2013). "Analysis of Solid and Aqueous Phase Products from Hydrothermal Carbonization of Whole and Lipid-Extracted Algae." *Energies*, Vol 7, Iss 1, Pp 62-79 (2013)(1): 62.

Discussion:

- The maximum lipid yield with sulfuric acid treatment was 18.28% at 115 °C for 25 minutes with 1.75 % sulfuric acid concentration.
- Highest lipid yield was achieved with liquid nitrogen treatment.